Generation of a Mixed-Valent Fe(III)Fe(IV) Form of Intermediate Q in the Reaction Cycle of Soluble Methane Monooxygenase, an Analog of Intermediate X in Ribonucleotide Reductase R2 Assembly

Ann M. Valentine,[†] Pedro Tavares,[‡] Alice S. Pereira,[‡] Roman Davydov,[§] Carsten Krebs,[‡] Brian M. Hoffman,[§] Dale E. Edmondson,^{||} Boi Hanh Huynh,[‡] and Stephen J. Lippard*,†

> Department of Chemistry Massachusetts Institute of Technology Cambridge, Massachusetts 02139 Department of Chemistry, Northwestern University Evanston, Illinois 60208 Departments of Biochemistry, Chemistry, and Physics Emory University, Atlanta, Georgia 30322

> > Received December 9, 1997

The selective conversion of methane to methanol under ambient conditions at the carboxylate-bridged diiron center in the hydroxylase component (MMOH) of soluble methane monooxygenase (sMMO) is a remarkable reaction. Several intermediates in the reaction cycle have been identified kinetically and spectroscopically, including diiron(III) peroxo (H_{peroxo}) and a formally dioxodiiron(IV) (Q) species.¹⁻³ These intermediates build up and decay following reaction of the fully reduced, diiron(II) form of MMOH with O₂. Similar chemistry occurs during the assembly of the structurally analogous dimetallic center in the R2 subunit of Escherichia coli ribonucleotide reductase (RNR), where an intermediate, mixed-valent state Fe(III)Fe(IV) species has been well characterized.⁴⁻⁶ The paramagnetic properties of intermediate X in R2 have been a valuable asset in its characterization, facilitating study by electron nuclear double resonance (ENDOR) spectroscopy.^{4,6} In the present paper, we present the results of rapid freeze quench (RFQ)/radiolytic reduction/Mössbauer spectroscopy which demonstrate that intermediate Q in the MMOH reaction cycle can be cryoscopically reduced to the paramagnetic Fe(III)Fe(IV) state. This species, which we designate Q_X , is closely related to intermediate X in the R2 assembly reaction. Its identification strengthens the spectroscopic link between the O₂ reaction chemistry of MMOH and R2 and paves the way for future ENDOR studies using isotopically labeled dioxygen for the structural characterization of Q.

Low-temperature radiolytic reduction has been applied to the study of carboxylate-bridged diiron proteins^{7,8} and models.⁹ In a solid matrix, a diamagnetic sample can be reduced by one electron to a paramagnetic species while maintaining a geometry close to that of the diamagnetic form. Annealing the sample to temperatures above 200 K allows the sample to relax to an equilibrium conformation, with concomitant conversion of the

- Feig, A. L.; Lippard, S. J. Chem. Rev. 1994, 94, 759-805.
 Liu, K. E.; Lippard, S. J. In Advances in Inorganic Chemistry; Sykes, A. G., Ed.; Academic Press, Inc.: San Diego, CA, 1995; Vol. 42, pp 263-
- (3) Wallar, B. J.; Lipscomb, J. D. Chem. Rev. 1996, 96, 2625-2657.
- (4) Strugeon, B. E.; Burdi, D.; Chen, S.; Huynh, B. H.; Edmondson, D.
 E.; Stubbe, J.; Hoffman, B. M. J. Am. Chem. Soc. 1996, 118, 7551–7557. (5) Riggs-Gelasco, P.; Shu, L.; Chen, S.; Burdi, D.; Huynh, B. H.; Que,
- (6) Willems, J.-P.; Lee, H.-I.; Burdi, D.; Doan, P. E.; Stubbe, J.; Hoffman,
- B. M. J. Am. Chem. Soc. 1997, 119, 9816-9824 (7) Davydov, R.; Kuprin, S.; Gräslund, A.; Ehrenberg, A. J. Am. Chem.
- Soc. 1994, 116, 11120-11128.
- (8) Davydov, A.; Davydov, R.; Gräslund, A.; Lipscomb, J. D.; Andersson,
 K. K. J. Biol. Chem. 1997, 272, 7022-7026.
 (9) Davydov, R. M.; Ménage, S.; Fontecave, M.; Gräslund, A.; Ehrenberg,
- A. J. Bioinorg. Chem. 1997, 2, 242-255.



Figure 1. Mössbauer spectra of a rapid freeze quench sample from the reaction of Hred/2B with dioxygen at 4 °C. The reaction was quenched at 6 s. Spectra were recorded at 4.2 K with a 50 mT magnetic field applied parallel to the Mössbauer low-energy γ rays. (A) The spectrum recorded before the sample was subjected to high-energy γ irradiation. The position of the doublet attributed to intermediate Q is indicated by a bracket. (B) The spectrum of the sample after high-energy γ irradiation. The signals due to the new paramagnetic species Q_X are designated by arrows. (C) A difference spectrum (B minus A) showing the conversion of a portion of the diamagnetic Q (upward pointing quadrupole doublet) into the paramagnetic Q_X (downward pointing absorption signals). The solid line is a theoretical simulation of Q using the previously reported parameters.¹¹

electron paramagnetic resonance (EPR) signal to that observed following chemical reduction in solution at 25 °C. This technique has thus far been applied only to the diferric form of non-heme iron proteins and model compounds, generating the mixed-valent Fe(II)Fe(III) form. Intermediate Q, which is diamagnetic at 4 K, has now been made paramagnetic in this manner.

Figure 1A depicts the Mössbauer spectrum of a RFQ sample containing intermediate Q (35% of total iron) as well as other protein species.^{10,11} The Mössbauer spectrum of the same sample after irradiation is shown in Figure 1B.¹² A difference spectrum (Figure 1C) illustrates that irradiation converts a portion of Q into a new paramagnetic species, designated Q_x. The amount of Q lost appears as a negative quadrupole doublet (pointing upward) and corresponds to approximately 22% of the total iron in the sample, while the new species accounting for the same portion

Massachusetts Institute of Technology.

[‡] Department of Physics, Emory University.

[§] Northwestern University.

Departments of Biochemistry and Chemistry, Emory University.

⁽¹⁰⁾ The sample was prepared with proteins from Methylococcus capsulatus (Bath) by rapid freeze quench (RFQ) techniques as reported previously.¹¹ An anaerobic solution of 0.7 mM MMOH_{red} and 1.4 mM MMOB in 25 mM MOPS (pH 7) was mixed rapidly at 4 °C with a dioxygen-saturated solution of 25 mM MOPS (pH 7) with 40% glycerol. The reaction mixture was allowed to age for 6 s before quenching into isopentane at -150 °C. The sample was packed into either a Mössbauer sample cell or a quartz Q band ENDOR tube for spectroscopic measurements. Analysis of the Mössbauer data indicates It is sample contained 32% H_{red} . 10% $H_{percons}$ 35% Q, 14% H_{ox} , and 9% V. Spectral parameters for these species have been reported previously¹¹ except for V, which is a new diamagnetic diiron(III) component having $\delta = 0.51$ in $\Delta E_Q = 2.34$ mm/s, parameters similar to those of the (μ -oxo)-diiron(III) form of R2.¹

⁽¹¹⁾ Liu, K. E.; Valentine, A. M.; Wang, D.; Huynh, B. H.; Edmondson, D. E.; Salifoglou, A.; Lippard, S. J. J. Am. Chem. Soc. 1995, 117, 10174-10185.

⁽¹²⁾ Samples were irradiated by using γ rays from a 60 Co source at a dose rate of 0.45 Mrad/h to a total dose of 6 Mrad.

Table 1. Mössbauer Parameters for Q_X and Related Species^a

	Qx		$R2-X^b$		$[Fe_2(6-Me-TPA)_2(\mu-O)_2]^{3+c}$	
	Fe _A	Fe _B	Fe _A	Fe _B	Fe _A	Fe _B
δ^d	0.48(6)	0.14(6)	0.56(3)	0.26(4)	0.48(3)	0.08(3)
ΔE_0^d	-0.9(1)	-0.6(1)	-0.9(1)	-0.6(1)	1.6(2)	0.5(1)
η	0.2(5)	0.9(5)	0.5(2)	2.7(3)	1.0(3)	1.0(3)
Á _{xx} ^e	-70(4)	26(4)	-74.2(2)	27.5(2)	-64.5(20)	20(3)
A_{vv}^{e}	-75(4)	30(4)	-72.2(2)	36.8(2)	-64.5(20)	36.5(15)
$A_{zz}^{j}e$	-75(4)	32(4)	-73.2(2)	36.8(2)	-64.5(20)	36.5(15)

^{*a*} Numbers in parentheses are estimated standard deviations in the last significant digits. ^{*b*} Data from ref 4. ^{*c*} Data from ref 14. 6-Me-TPA = *N*-(6-methyl-2-pyridylmethyl)-*N*,*N*-bis(2-pyridylmethyl)amine. ^{*d*} Units mm/s. ^{*e*} Units MHz.



Figure 2. Mössbauer spectral comparison of Q_X and R2-X. (A) 4.2 K spectrum of Q_X prepared from the spectrum in Figure 1B by removing the contributions of H_{red} (40% of total iron absorption), H_{peroxo} (9.5%), Q (13.5%), V (8.5%),¹⁰ and H_{ox} (6.5%). The solid line plotted through the experimental data is a theoretical simulation of Q_X obtained by using the parameters listed in Table 1. The corresponding theoretical spectrum of R2-X⁴ is shown in B. (C) A difference spectrum (parallel minus perpendicular) showing the field direction dependence of Q_X . The solid line plotted through the experimental data is a theoretical simulation of this dependence obtained by using the parameters listed in Table 1. The corresponding difference spectrum for the R2-X intermediate⁴ is shown in D. The positions for the two sets of $\Delta m = 0$ lines corresponding to the two Fe sites are indicated by brackets at the bottom of the figure.

appears as a magnetic spectrum in the positive direction (pointing downward). Detailed analysis of the spectra indicates that high-valent Q was reduced preferentially over other species during irradiation, except for a small portion (7.5% of the total iron) of H_{ox} which was apparently fully reduced to the differous form.

Because the Mössbauer parameters of the other components are known,^{11,13} their contributions can be removed from the spectrum of the irradiated sample to reveal the features of Q_X (Figure 2A). Its properties are very similar to those of R2-X (Figure 2B), formally an Fe(III)Fe(IV) species.⁴ The close resemblance of the electronic structures of Q_X and R2-X is further revealed by the field direction dependence of their Mössbauer spectra. For an isotropic $S = \frac{1}{2}$ system, such as the exchangecoupled Fe(III)Fe(IV) state of R2-X, the weak-field spectrum depends strongly on the direction of the applied field in relation to the direction of the Mössbauer γ rays. The $\Delta m = 0$ transition lines acquire their minimum and maximum intensities, respectively, when the field is oriented in a parallel and perpendicular direction. In the irradiated sample, all species except for Q_X are either diamagnetic or have an integer spin, and their spectra do not change with the direction of a weak applied field. Consequently, a difference spectrum (Figure 2C) recorded with a 50 mT field applied parallel and perpendicular to the γ rays will cancel the contributions from all other species, revealing the field direction dependence of Q_X . Two sets of $\Delta m = 0$ lines (marked by brackets) corresponding to two distinct Fe sites are clearly visible. This difference spectrum is very similar to the analogous one of R2-X (Figure 2D).⁴ Preliminary analysis of these Q_X spectra yields Mössbauer parameters comparable to those reported for R2-X and a related model compound (Table 1).^{4,14}

The structure of intermediate Q has been inferred from a variety of physical and theoretical studies,^{11,13,15-18} but owing in part to its diamagnetism is less well defined than that of intermediate X of RNR-R2. The rhombic nature of the EPR spectrum of R2-X at 35 GHz (Q band) has been exploited by ENDOR spectroscopy to afford a very detailed picture of the structure, as well as the fate of oxygen atoms incorporated during its formation from reduced R2 and O₂.^{4,6} Such studies on Q would prove invaluable in its characterization. The 35 GHz EPR spectrum of a RFO sample of Q quenched into a sample tube and then irradiated exhibits an intense signal at g = 2 due to free radicals generated by the high energy γ rays (data not shown). This signal completely obscures the Q_X EPR features, as would be expected if its dispersion from g = 2 were as small as that of R2-X.¹⁹ Efforts are underway to reduce the intensity of the free radical signal so that the EPR spectrum of Q_x may be revealed.

In conclusion, this work, together with recent experiments on wild-type and mutant forms of R2,²⁰ demonstrates the first spectroscopic link between sMMO intermediate Q and RNR R2 intermediate X. As such it provides strong support for the supposition^{1,21,22} that the reaction chemistry of the diiron centers in the two proteins is quite analogous. The present application of the radiolytic reduction technique extends its utility to high-valent iron species and facilitates detailed ENDOR studies of intermediate Q. Knowledge of the structure of Q is crucial for understanding the sMMO reaction cycle.

Acknowledgment. This work was supported by grants from the National Institutes of Health (GM32134 to S.J.L., GM47295 to B.H.H. and D.E.E., and HL13531 to B.M.H.). P.T. and A.S.P. are recipients of postdoctoral research fellowships from Programa PRAXIS XXI of Junta Nacional de Investigacao Cientifica e Tecnologica, Portugal. We thank Professor JoAnne Stubbe for the use of RFQ equipment for the preparation of EPR samples and Drs. Pamela Riggs-Gelasco and Douglas Burdi for technical assistance.

JA974169X

- (14) Dong, Y.; Que, L., Jr.; Kauffmann, K.; Münck, E. J. Am. Chem. Soc. **1995**, *117*, 11377–11378.
- (15) Lee, S.-K.; Fox, B. G.; Froland, W. A.; Lipscomb, J. D.; Münck, E. J. Am. Chem. Soc. **1993**, 115, 6450–6451.

(16) Lee, S.-K.; Nesheim, J. C.; Lipscomb, J. D. J. Biol. Chem. 1993, 268, 21569-21577.

(17) Shu, L.; Nesheim, J. C.; Kauffmann, K.; Münck, E.; Lipscomb, J. D.; Que, L., Jr. *Science* **1997**, *275*, 515–518.

(18) Siegbahn, P. E. M.; Crabtree, R. H. J. Am. Chem. Soc. 1997, 119, 3103-3113.

(19) The EPR spectrum also exhibits a signal with $g \le 1.96$ corresponding to less than 2% of the total iron. This feature arises from a mixed valent Fe(II)Fe(III) species, which is not present in a sufficient population to be detected by Mössbauer spectroscopy.

detected by Mossbauer spectroscopy.
(20) (a) Tong, W. H.; Chen, S.; Lloyd, S. G.; Edmondson, D. E.; Huynh, B. H.; Stubbe, J. J. Am. Chem. Soc. 1996, 118, 2107–2108. (b) Bollinger, J. M.; Krebs, C.; Vicol, A.; Chen, S.; Ley, B. A.; Edmondson, D. E.; Huynh, B.

H. J. Am. Chem. Soc. In press. (21) Nordlund, P.; Eklund, H. Current Opin. Struct. Biol. 1995, 5, 758– 766.

(22) Que, L., Jr.; Dong, Y. Acc. Chem. Res. 1996, 29, 190-196.

⁽¹³⁾ Liu, K. E.; Wang, D.; Huynh, B. H.; Edmondson, D. E.; Salifoglou, A.; Lippard, S. J. J. Am. Chem. Soc. **1994**, 116, 7465–7466.